



Genome sequence of *Pseudomonas aeruginosa* strain DK1-NH57388A, a stable mucoid cystic fibrosis isolate

Norman, Anders; Ciofu, Oana; Amador, Cristina Isabel; Høiby, Niels; Jelsbak, Lars

Published in:
Genome Announcements

DOI:
[10.1128/genomeA.00008-16](https://doi.org/10.1128/genomeA.00008-16)

Publication date:
2016

Document version
Publisher's PDF, also known as Version of record

Document license:
[CC BY](#)

Citation for published version (APA):
Norman, A., Ciofu, O., Amador, C. I., Høiby, N., & Jelsbak, L. (2016). Genome sequence of *Pseudomonas aeruginosa* strain DK1-NH57388A, a stable mucoid cystic fibrosis isolate. *Genome Announcements*, 4(1), [e00008-16]. <https://doi.org/10.1128/genomeA.00008-16>

Genome Sequence of *Pseudomonas aeruginosa* Strain DK1-NH57388A, a Stable Mucoïd Cystic Fibrosis Isolate

Anders Norman,^a Oana Ciofu,^b Cristina Isabel Amador,^a Niels Høiby,^{b,c} Lars Jelsbak^a

Technical University of Denmark, Department of Systems Biology, Kongens Lyngby, Denmark^a; Department of Immunology and Microbiology, Costerton Biofilm Center, University of Copenhagen, Copenhagen, Denmark^b; Department of Clinical Microbiology, Rigshospitalet, Copenhagen, Denmark^c

***Pseudomonas aeruginosa* is an important opportunistic pathogen associated with chronic pulmonary infections and mortality in cystic fibrosis (CF) patients. Here, we present the complete genome sequence of stable mucoïd *P. aeruginosa* strain DK1-NH57388A, a CF isolate which has previously been used to establish chronic lung infections in an animal model.**

Received 5 January 2016 Accepted 6 January 2016 Published 25 February 2016

Citation Norman A, Ciofu O, Amador CI, Høiby N, Jelsbak L. 2016. Genome sequence of *Pseudomonas aeruginosa* strain DK1-NH57388A, a stable mucoïd cystic fibrosis isolate. Genome Announc 4(1):e00008-16. doi:10.1128/genomeA.00008-16.

Copyright © 2016 Norman et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Lars Jelsbak, lj@bio.dtu.dk.

The Gram-negative bacterium *Pseudomonas aeruginosa* DK1-NH57388A is a stable mucoïd isolate, collected in 1997 from the sputum of a Danish cystic fibrosis (CF) patient (CF86) who has had chronic lung infection since 1983. DK1-NH57388A is a representative isolate from the *P. aeruginosa* DK1 lineage which is highly prevalent in Danish CF patients (1, 2). The alginate hyper-producing phenotype of DK1-NH57388A is caused by disruption of the *mucA* anti-sigma factor gene. Furthermore, it has a functional *N*-acyl-homoserine lactone (AHL)-based quorum-sensing (QS) system and produces QS regulated exoproducts including elastase, pyocyanin, and chitinase (3, 4). Due to the stability of the alginate phenotype during several passages, the isolate has been used in an animal model of chronic lung infection (3). This animal model has been used to test the effects of azithromycin, novispirin, cysteamine, oligo(G), and bacteriophages on infection (4–8).

Genomic DK1-NH57388A DNA was isolated using the Wizard genomic DNA purification kit (Promega) according to the manufacturer's instructions. Genome sequencing was performed on the Illumina MiSeq platform, resulting in 2,797,134 raw 150-bp paired-end reads with a median insert size of 410 bp. Reads were first screened for typical contaminants by using BWA (9) to map against known sequences such as the phiX control phage, leading to the removal of 25,743 reads (0.9%). The SPAdes genome assembler v3.6.2 (10) was then used to *de novo* assemble 417,369,767 quality trimmed (11) bases, resulting in 53 scaffolds (N_{50} : 351 Kbp, median \times coverage: 66.7). A complete draft genome was then constructed in Geneious version R8.3 (12) using the genome sequence of the closely related *P. aeruginosa* DK2 strain (GenBank accession no. CP003149) as a guide. Raw sequencing reads were then mapped back to the draft genome sequence in order to resolve single-nucleotide polymorphisms in repetitive regions and to close gaps in poorly covered regions. The final genome sequence comprises a single circular chromosome, 6,212,531 bp in length, with an average G+C content of 66.6%. Genome size and G+C content are both consistent with other previously sequenced *P. aeruginosa* strains. Automatic genome annotation was performed using Prokka resulting in 5,632 coding regions (CDS), 62

tRNA genes, 12 rRNA genes, and a single transfer-messenger RNA (tmRNA) gene. One of the four rDNA regions contain a disrupted 16S gene due to a 261-bp deletion and a disrupted 23S gene, due to the insertion of the 1,236-bp mobile element IS222. Furthermore, the *mucA* gene has been disrupted by a 105-bp deletion as previously described (3).

The DK1-NH57388A genome is 190 Kbp smaller than the previously described transmissible DK2 strain (6,402,658 bp) which is also prevalent in Danish CF patients (1). The two genomes share 5,988,592 identical sites (90.7%) as revealed by multiple genome alignment using Mauve. Furthermore, DK1 contains two clusters of regularly interspaced short palindromic repeat (CRISPR) systems, one identical to the one found in DK2 (genomic island 5) and one with >95% nucleotide identity to an 11 Kbp CRISPR-cassette found in *P. aeruginosa* RP73 (13). The DK1-NH57388A genome will enable epidemiological studies of the DK1 lineage.

Nucleotide sequence accession number. The complete DK1-NH57388A genome sequence has been deposited in the ENA under the accession no. [LN870292](https://ena.ebi.ac.uk/ena/record/LN870292/).

FUNDING INFORMATION

Villum Fonden (Villum Foundation) provided funding to Lars Jelsbak under grant number VKR023113.

REFERENCES

- Jelsbak L, Johansen HK, Frost A-L, Thøgersen R, Thomsen LE, Ciofu O, Yang L, Haagenen JAJ, Høiby N, Molin S. 2007. Molecular epidemiology and dynamics of *Pseudomonas aeruginosa* populations in lungs of cystic fibrosis patients. Infect Immun 75:2214–2224. [http://dx.doi.org/10.1128/IAI.01282-06](https://doi.org/10.1128/IAI.01282-06).
- Marvig RL, Damkjaer S, Khademi SMH, Markussen TM, Molin S, Jelsbak L. 2014. Within-host evolution of *Pseudomonas aeruginosa* reveals adaptation toward iron acquisition from hemoglobin. MBio 5:e00966-14. [http://dx.doi.org/10.1128/mBio.00966-14](https://doi.org/10.1128/mBio.00966-14).
- Hoffmann N, Rasmussen TB, Jensen PØ, Stub C, Hentzer M, Molin S, Ciofu O, Givskov M, Johansen HK, Høiby N. 2005. Novel mouse model of chronic *Pseudomonas aeruginosa* lung infection mimicking cystic fibrosis. Infect Immun 73:2504–2514. [http://dx.doi.org/10.1128/IAI.73.4.2504-2514.2005](https://doi.org/10.1128/IAI.73.4.2504-2514.2005).
- Hoffmann N, Lee B, Hentzer M, Rasmussen TB, Song Z, Johansen HK,

- Givskov M, Høiby N. 2007. Azithromycin blocks quorum sensing and alginate polymer formation and increases the sensitivity to serum and stationary-growth-phase killing of *Pseudomonas aeruginosa* and attenuates chronic *P. aeruginosa* lung infection in *Cftr*^{-/-} mice. *Antimicrob Agents Chemother* 51:3677–3687. <http://dx.doi.org/10.1128/AAC.01011-06>.
5. Hengzhuang W, Song Z, Ciofu O, Onøyen E, Rye P, Høiby N. OligoG CF-5/20 disruption of mucoid *Pseudomonas aeruginosa* biofilm in a murine lung infection model. *Antimicrob Agents Chemother*, in press.
 6. Song Z, Wu H, Mygind P, Raventos D, Sonksen C, Kristensen H-H, Høiby N. 2005. Effects of intratracheal administration of novispirin G10 on a rat model of mucoid *Pseudomonas aeruginosa* lung infection. *Antimicrob Agents Chemother* 49:3868–3874. <http://dx.doi.org/10.1128/AAC.49.9.3868-3874.2005>.
 7. Charrier C, Rodger C, Robertson J, Kowalczyk A, Shand N, Fraser-Pitt D, Mercer D, O'Neil D. 2014. Cysteamine (Lynovex®), a novel mucoactive antimicrobial & antibiofilm agent for the treatment of cystic fibrosis. *Orphanet J Rare Dis* 9:189.
 8. Alemayehu D, Casey PG, McAuliffe O, Guinane CM, Martin JG, Shanahan F, Coffey A, Ross RP, Hill C. 2012. Bacteriophages MR299-2 and NH-4 can eliminate *Pseudomonas aeruginosa* in the murine lung and on cystic fibrosis lung airway cells. *MBio* 3:e00029-12. <http://dx.doi.org/10.1128/mBio.00029-12>.
 9. Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics* 25:1754–1760. <http://dx.doi.org/10.1093/bioinformatics/btp324>.
 10. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
 11. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <http://dx.doi.org/10.1093/bioinformatics/btu170>.
 12. Kears M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. 2012. Geneious basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28:1647–1649. <http://dx.doi.org/10.1093/bioinformatics/bts199>.
 13. Jeukens J, Boyle B, Bianconi I, Kukavica-Ibrulj I, Tümmler B, Bragonzi A, Levesque RC. 2013. Complete genome sequence of persistent cystic fibrosis isolate *Pseudomonas aeruginosa* strain RP73. *Genome Announc* 1(4):e00568-13. <http://dx.doi.org/10.1128/genomeA.00568-13>.